Website: https://ijmpr.in/ | Print ISSN: 2958-3675 | Online ISSN: 2958-3683

NLM ID: 9918523075206676

Volume: 4, Special Issue:3 (May-June 2023); Page No: 83-94



An Updated Review (Phytochemistry, Antimicrobial Pharmacology) on Indigenous King of Bitter (*Andrographis Paniculata*)

Navneet Kumar Verma^{*1}, Prashant Singh², Sushil Kumar Tiwari¹, Shiwani Jaiswal¹, Karunakar Prasad Dwivedi¹, Vinay Kumar³, Shreya Maddheshiya³

- Associate Professor, Buddha Institute of Pharmacy, GIDA, Gorakhpur, UP, India-273209
- ² Professor, Buddha Institute of Pharmacy, GIDA, Gorakhpur, UP, India-273209
- ³ Assistant Professor, Buddha Institute of Pharmacy, GIDA, Gorakhpur, UP, India-273209

ABSTRACT

Andrographis paniculata (King of Bitters), also known as Kalmegh, is a member of the Acanthaceae family. Andrographis paniculata is widely cultivated, and its relevance as a medicinal plant is expanding as evidence of its several therapeutic applications grows. Considering the plant's beneficial properties, it might be recommended as a safe and vital medicinal herb for mankind. This herb is rich in chemical components, including lactones, diterpenoids, diterpene glycosides, flavonoids, and flavonoid glycosides. It possesses numerous pharmacological effects, including antibacterial, hepatoprotective, anti-cancer, anticancer, hypoglycemic, immunomodulatory, and hypotensive action. The goal of this study is to conduct a literature review on Andrographis paniculata, specifically papers on therapeutic advantages, chemical features, and pharmacological evaluation. It is widely utilised as a home cure for numerous maladies in the Bangladeshi traditional system of medicine. Steroids, phenols, terpenoids, alkaloids, saponins, and flavonoids were the active chemicals found in Andrographis paniculata. The presence of active components, andrographolide and neoandrographolide, which are diterpenoids derivatives, contributes to the therapeutic potential of this plant.

Key Words: Andrographis paniculata, Kalmegh, Andrographolide, King of Bitters.



*Corresponding Author

Navneet Kumar Verma

Associate Professor, Buddha Institute of Pharmacy, GIDA, Gorakhpur, UP, India-273209

INTRODUCTION

Andrographis paniculata is a branching annual with lanceolate green leaves that grows to a height of 60-70 cm [1]. 2. It thrives in Asian countries such as India, Sri Lanka, Pakistan, Java, Malaysia, and Indonesia. It is generally known as Kalmegh in India and is primarily found in the country's plains. It is one of the most commonly used medicinal plants in Ayurvedic and Unani systems of medicine. The plant is sometimes referred to as the 'King of Bitters' [2-3].

Because it has an exceptionally bitter taste in all parts of the plant. According to a literature review, the aerial portions of plants (leaves and stems) are most typically employed to extract active phytochemicals; however, the whole plant or mentioned to a limited extend [3]. A. paniculata has a broad spectrum of pharmacological effects and some of them are extremely beneficial such as Hepatoprotective, antimicrobial, antifungal, antioxidant, anti-inflammatory, antipyretic, anticancer and antidiarrhoeal effects. According to Unani system of medicine it is useful in the treatment of chronic hepatitis.



Figure.1; Andrographis paniculata Plant

PHYTOCHEMICAL CONSTITUENTS

A review of the literature reveals that the presence of various chemical constituents in the aerial parts of the Andrographis paniculata are andrographolide, which is diterpene lactone, colourless, crystalline, bitter in taste [4]. Other compounds 14-deoxy-11-oxoandrographolide, didehydro andrographolide/andrographlide 14deoxyandrographolide, non-bitter compound is neo andrographolide, homoandrographolide, andrographosterin, andrograpanin, α- sitosterol, stigmasterol. Apigenin-7, 4-dio-methyl ether, 5-hydroxy 7,8,2, 3-tetramethoxy flavones, monohydroxy trimethyl flavones, andrographin, dihydroxy di-methoxy flavoue, panicolin, andrographoneo, andrographoside, andropani-culoside A(3,7,8) andrograpanin, soandrographolide and skollcaflavone (912). Six 3-o-beta-Dglucopyranosyl-14, 19-dideoxyandrographolide, entlabdane diterpenoids i.e. 14-deox, hydroxyandrographolide, 19-o-[beta-D-apiofuranosy 1-2beta-D-glucopyranoyl]-3, 14-dideoxyandiographolide, 3-obeta-Dglucopyranosyl- andro-grapholide,12S-hydroxy andrographolide and andrographatoside. These compounds showed inhibitor activity against several fungal and bacterial strains. Dua et al. reported four xanthones 1,8-dihydroxy3,7dimethoxy xanthone, 4,8-di-hydroxy-2, 7-dimethoxyxanthones, 1,2-dihydroxy-6, 8-dimethoxyxanthoneand 3,7,8trimethoxy-1-hydroxyxanthone from the roots [5].

Andrographolide Neoandrographolide 14-Deoxy-11,12didehydroandrographolide

Furthermore, antibiotics usage causes some common side effects, including hypersensitivity and depletion of beneficial gut microorganism [6,7]. Acute upper respiratory tract infections (URTIs) is another significant cause of antibiotic resistance since physicians needlessly write antibiotic prescriptions [8-12]. Even though the vast majority of URTIs are mild, self-diagnosed and self-treated, they are the most common reason for absenteeism from school or work [13]. URTIs can be mainly caused by viruses, such as a rhinovirus, influenza virus, adenovirus, enterovirus, and respiratory syncytial virus. Bacteria like S. pyogenes, a Group A Streptococcus, may cause roughly 15% of sudden onset pharyngitis presentations [8]. Viral pharyngitis is mainly treated based on the symptoms that appeared, whereas bacterial pharyngitis can be treated with antibiotics. However, current evidence does not support the usefulness of antibiotics treatment in non-specific URTIs [13,14]. Therefore, research is urgently required to find alternatives to conventional medications for eradicating IDs. Natural products based therapy could be an excellent source of antimicrobial agents that would offer symptomatic relief since they have the high potentiality to inhibit the growth of microbes in the host-defence mechanism [4], as well as they offer promising outcomes in the scientific investigations [15-20]. Additionally, it would reduce unnecessary antibiotic prescription; therefore, the chances of antibiotic-resistance would be reduced. Nowadays, the philosophy of drug discovery has transformed into "one drug, multitarget" from "one drug, one target" [16.21–28]. Plant-derived secondary metabolites hold the potential of multi-targeting properties as they need to undergo evolving defence mechanisms of the plant against predators like bacteria, fungi, virus, even insects and herbivores [15,16,21,22,29,30]. A majority of the world population relies on medicinal plants for first-line treatment due to the severe side effects of synthetic drugs [31]. Moreover, plants' ability to cure diseases and the necessity of their study in sacred texts motivated people to use natural remedies and researchers to study their pharmacology [32,33]. Plant-based secondary metabolites commonly isolated are phenols, tannins, flavonoids, lignans, terpenes, and a wide range of alkaloids [21]. Since natural products are better models with ideal pharmacokinetics/ pharmacodynamics properties [16], often feature biologically relevant molecular scaffolds and pharmacophore patterns that have evolved as preferred ligandprotein binding motif [22], they gained tremendous importance for the development of polypharmacological drugs for IDs, cancers, and neurological disorders [22,34]. Furthermore, about 80% of drugs are either natural products or analogues mimicking them, and steadily increasing approval rate (after the 1990s, the average annual approval rate is 10.3) of natural product-derived drugs from the US Food and Drug Administration (FDA) have encouraged researchers and pharmaceutical industries to search the effective multitarget drugs for various ailments [30,34]. Currently, a number of natural products, including morphine, quinine, reserpine, cocaine, and ephedrine, are now available in pure form as drug substances [15]. Besides, many pure compounds are identified by pharmaceutical scientists worldwide because of having advanced technology that eases and fasten the characterization and structural elucidation of isolated metabolites. Capitalizing on these findings are crucial for medical advancement to overcome unavoidable circumstances happed by

synthetic drugs. One attractive medicinal plant and its metabolites that have gained considerable and progressive interest for decades are Andrographis paniculata (Burm. f.) Wall. ex Nees. This annual plant belongs to the Acanthaceae family and is commonly known as "King of thebitters" or "Kalmegh". It is native to India and Sri Lanka and widely found in Southern and Southeastern Asia, including Bangladesh, China, Hong Kong, Indonesia, Malaysia, Myanmar, Philippines, and Thailand [35–39]. Usually, the aerial parts, roots or leaves of A. paniculata are used separately. These plant parts are used traditionally as powder, infusion, or decoction form either alone or in combination with other medicinal plants for the treatment of leprosy, gonorrhoea, respiratory tract infections, scabies, boils, skin eruptions, chronic and seasonal fevers, griping, irregular bowel habits, loss of appetite, alopecia, general debility, diabetes, jaundice, dyspepsia, hemopathy, cough, oedema, liver complaints, dysentery, malaria, enteritis, helminthiasis, herpes, peptic ulcer, skin infections (topical use), and snake-bites (topical use) [35,36,39,40]. Modern science is focusing on validating the traditional claims of this plant through systemic investigations. Different extracts (i.e., acetone, chloroform, ethanol, hexane, methanolic or aqueous extract) and isolated pure metabolites from A. paniculata have been investigated for properties, example, antibacterial, antifungal, for antiviral, antiparasitic, hypocholesterolemia, anti-inflammatory, anti-hyperglycemic, hepatoprotective, anticancer, immunomodulatory, cardiovascular, antihyperlipidemic, emollient, anti-snake venom, anti-platelet aggregation, anti-fertility, carminative, and antipyretic properties at in vitro and in vivo conditions [41–53]. Clinical trials were conducted to validate the in vitro and preclinical antimicrobial pharmacology of A. paniculata. Several clinical trials have justified the anti-infectious activity of A. paniculata against URTIs, influenza, and HIV as well as its effectiveness in treating osteoarthritis and multiple sclerosis [54-59]. The clinical studies conducted using the standardized A. paniculata extracts alone or in combination with other medicinal plants (i.e., Kan Jang, Kalm ColdTM). The extracts reported having approximately 4-6% of andrographolide [56,60], responsible for the beneficial effects of the plant extracts. In recent years, there are few more clinical studies conducted [12,61-73]. However, to date, Andrographolide or A. paniculata extracts have not achieved the human clinical trial stage. This might be because of low bioavailability that is attributed to the fast biotransformation and removal from the body [74] or due to a lack of a well-defined mechanism of action because it is considered as a highly promiscuous compound and engaged in covalent interactions with numerous previously unknown cellular targets in cell type-specific manner [75]. There are two important and typical challenges: (i) poor pharmacokinetics and (ii) limited bioavailability are involved with many natural products, for example, sitosterol, quercetin, genistein, rutin etc., prevent their in vitro to preclinical or clinical translation [76]. Nevertheless, there is a numerous investigation reported to have potential efficacy of A. paniculata for IDs as well as other ailments, including clinical efficacy. So, would A. paniculata extracts and their metabolites be a choice of better therapeutics? Would it be undergone for clinical development? Hence, there is a need to explore further the feasibility of A. paniculata with potential use as anti-infective agents for a wide range of invasive microbes. A better understanding of the mode of actions of the active ingredients of A. paniculata would enhance the clinical development of this drug. Additionally, there were several recent in vitro, in vivo, and a few clinical trials reported its potential therapeutic efficacy for antimicrobial pharmacology. To our best knowledge, critical evaluation of A. paniculata's purported benefits is not substantial yet; hence, it would be important to review them before conducting further research in this area.

PHARMACOLOGICAL ACTIVITIES:

The usage of various sections of A. paniculata in folk medicine prompted scientists to investigate its pharmacological properties in order to validate this plant as a medicinal agent. Many studies have shown that this plant has antimicrobial, cytotoxic, antiprotozoan, antiinflammatory, antioxidant, immunostimulant, antidiabetic, antiinfective, hepatorenal protective, sex hormone modulatory, liver enzymes modulatory, insecticidal, neuroprotective, anticancer, antipyretic, antiplatelet, and toxicity properties [77-79].

Anti-microbial activity:

Aqueous extract, andrographolide and arabinogalactan proteins that were isolated from the dried herb of *A. paniculata* were screened for anti-microbial activity. The results showed that aqueous extract and arabinogalactan proteins possess antibacterial activity against *Bacillus subtilis* (*B. subtilis*), *Escherichia coli* (*E.coli*), *Pseudomonas aeruginosa* while andrographolide was the only one which is active against *B. subtilis*. All three were also reported to possess antifungal activity against *Candida albicans* [77]. The five rare noriridoides, andrographidoides (A-E) were screened for their anti-bacterial activity against *E. coli*, *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Pseudomonas aeruginosa* and *B. subtilis*. None of the compounds showed inhibitory activity (MIC>100 μg/mL). Gentamycin, chloramphenicol and ciprofloxacin were used as positive controls [80].

Anti-inflammatory/Anti-allergic activity:

Aqueous extract when combined with methanol leaves extract showed significant alleviation of lipopolysaccharides that induced release of pro-inflammatory (NO, IL-1 β and IL-6), inflammatory (PGE2 and TXB2) and allergic mediators (LTB4). No inhibition was observed against histamine release (Chandrasekaran *et al.* 2010). Seven photochemicals, namely, andrographolide, neoandrographolide, isoandrographolide, andrograpanin, 7-O-methylwogonin, 14-deoxy-11,12- didehydroandrographolide and skullcapflavone (Figure 1) isolated from *A. paniculata* leaves were screened for *in vitro* anti-inflammatory and anti-allergic potential. The results showed that andrographolide, isoandrographolide, 7-Omethylwogonin and skullcapflavone-1 significantly inhibited inflammatorymediators NO and PGE2 release from

lipopolysacharide (LPS) stimulated cultured macrophages. However, IL-1β production in LPS stimulated macrophages was inhibited by andrographolide, isoandrographolide and 7-O-methylwogonin. Also, IL-6 production from LPS induced macrophages was significantly (P<0.01) inhibited by andrographolide, isoandrographolide and skullcapflavone-1 in a concentration dependent manner. The results also showed that andrographolide, isoandrographolide and skullcapflavone-1 significantly suppressed TXB4 released in A23187 activated HL-60 promyelocytic leukemia cells. Furthermore, the anti-allergic properties of the phytoconstituents was investigated on A23187 induced LTB4 production. The result showed 30.5% and 19.6% inhibition of LTB4 production in A23187 induced HL-60 promyeolocytic leukemia cells at concentrations of 63 µM and 33.5 µM for skullcapflavone and 7-O-methylwogonin respectively. The IC50 value for the reference standard captopril was 48 µmol/L. 7-O-methylwogonin was the only phytoconstituent that potently inhibited A23187 induced histamine release in RBL-2H3 rat basophil leukemic cells in a dose dependent manner[46]. Andrographolide, dehydroandrographolide and neoandrographolide isolated from the aerial parts of A. paniculata exhibited anti-inflammatory effects by interfering with COX enzyme activity. Andrographolide (30.1 µM) and dehydroandrographolide (28.5 µM) markedly inhibited COX-1 in ionophore A23187-induced human platelets. Dehydroandrographolide (28.5 µmol/L) and neoandrographolide (20.8 µM) strongly suppressed the LPS-stimulated COX-2 activity in human blood. In addition, dehydroandrographolide modulated the level of LPS-induced TNF- α , IL-6, IL-1β, and IL-10 secretion in human blood in a concentration dependent manner, showing that dehydroandrographolide has the highest efficacy. The result further showed that the mechanism of dehydroandrographolide may be related to down-expression of genes involved in the inflammatory cascade [81].

Andrograpanin (15-90 μ M) isolated from the ethanol extract of the leaves inhibited NO and pro-inflammatory cytokines (TNF α , IL-6, IL-12p70) in a dose dependent manner from lipopolysaccharide activated macrophages. Significant (P<0.05) inhibition of NO was evident at a concentration of 30 μ mol/L and at a concentration of 75 μ M. Andrograpanin almost completely inhibited NO production. Significant inhibition of pro-inflammatory cytokines was evident at a concentration of 1.5 μ mol/L and there was an almost complete inhibition at a concentration of 90 μ mol/L. The RT-PCR and western blotting assays showed that andrograpanin inhibited productions of NO and pro-inflammatory cytokines through down-regulating iNOS and pro-inflammatory cytokines gene expression levels as well as p38 mitogen activator kinase signaling pathways. Further study showed that andrograpanin has more ability of down regulating IL-12 p35 and p40 proteins than their mRNA levels. This suggests that andrograpanin might be involved in down regulating the post-translation of IL-12 p35 and 40 proteins [82].

Antioxidant activity:

Andrographolide and aqueous extract of A. paniculata were screened for their antioxidant activity on nicotine induced oxidative stress in liver, kidney, heart, lungs and spleen of male wistar rats. The results showed that intraperitoneal administration of A. paniculata (25 mg/kg) and Aphanamixis polystachya (25 mg/kg) for a period of 7 days significantly (P<0.05) reduce levels of lipid peroxidation and increase antioxidant enzymes status in five organs screened as compared to nicotine [83]. The methanolic and aqueous leaves extracts of A. paniculata, andrographolide and 14-deoxy-11, 12-didehydroandrographolide exhibited lipid peroxidation inhibition in Srague Dawley rats and free radical scavenging activity against Diphenyl picrylhydrazyl (DPPH). The lipid peroxidation inhibition activity varied from 55.6% to 63.9% and 33.78% to 33.77% for methanolic and water extracts respectively. The activity of the methanolic extracts were higher and significantly different (P<0.05) from that of the water extract. The methanolic extract exhibited free radical scavenging activity ranging from 45.67% to 53.82%. The activity of andrographolide was 40.2% and for 12didehydroandrographolide it was 46.43%. The water extract exhibited poor free radical scavenging activity ranging from 25.29% to 28.77%. The methanolic, water extracts and isolated compounds exhibited a lower free radical scavenging activity as compared to quercetin (89%) and butylated hydroxylanisole (71%) used as positive controls [84]. A fourteenday oral treatment of Sprague Dawley rats with methanolic extract (1g/kg body weight) of dried leaves followed by carbon tetrachloride (CCl4) challenge preserved antioxidant enzyme-catalase and superoxide dismutase activities in erythrocytes. Whereas, lipid peroxidation, alanine transaminase, aspartate transaminase and plasma thiobarbituric acid reactive substances were restored to values as compared to that who did not receive CCl4. Andrographolide, 14-deoxy-11, 12- didehydroandrographolide were traceable in rat plasma followed by an oral dose of methanolic dried leaves extract (1 g/kg body weight) suggesting that these diterpenes may be responsible for the observed antioxidant activity [85].

Antiprotozoan activity:

Four xanthones were isolated from root fractions and screened for antiplasmodial activity against *Plasmodium falciparum*. Only a compound 1,2-dihydroxy-6,8-dimethoxyxantone possessed substantial antiplasmodial activity against *Plasmodium falciparum* with an IC50 value of 4µg/ ml. This compound also exhibited *in vivo* antimalarial activity in mice infected with *Plasmodium berghei* where it produced substantial reduction (62%) in parasitemia [42]. This study involved the root fractions that showed higher antimalarial activity ascompared to a previous study of fractions isolated from the leaves [86]. Andrographolide, neoandrographolide, deoxyandrographolide and andrographoside isolated from the leaves have been shown to possess some activity against *Plasmodium berghei* NK65 in *Mastomys natalensis* [87].

Insecticidal activity:

The ovicidal and larvicidal activity of the crude leaves extract of *A. paniculata* with five different solvents like benzene, hexane, ethylacetate, methanol and chloroform were tested against early third instar larvae of *Culex quinquefasciatus* (Say) and *Aedes aegypti* (Linn). The benzene, hexane, ethylacetate, methanol and chloroform extracts were found to be more effective against *Culex quinquefasciatus* than *Aedes aegypti*. The LC50 values were 112.19, 137.48, 118.67, 102.05, 91.20 mg/L and 119.58, 146.34, 124.24, 110.12, 99.54 mg/L respectively. The methanol and ethyl acetate extracts were found to be most effective for ovicidal activity against the two mosquito species. The extract of methanol and ethylacetate also exerted 100% mortality at a concentration of 200mg/L against *Culex quinquefasciatus* and at 250 mg/L against *Aedes aegypti* [88].

Antiinfective activity:

The efficacy of the leaves extract of *A. paniculata* in the treatment of symptoms of uncomplicated upper respiratory tract infection has been reported. The findings obtained in a randomized double blind placebo controlled clinical evaluation using the visual analogue scale for quantification of symptoms. It showed that the Kalmcold treatment significantly (P<0.05) decreased all the symptoms score except for earache whereas symptoms remained unchanged or got worse after Day 3 for the placebo group. The study revealed that Kalmcold was 2.1 times or 52.7% more effective than placebo in reducing symptoms of uncomplicated upper respiratory tract infection [89]. *A. paniculata* extract SHA-10 (1200 mg/day) administered for a period of five days. It significantly (P<0.05) reduced the intensity of the symptoms (tiredness, sleeplessness, sore throat and nasal secretion) in uncomplicated common cold at the beginning of Day 2 treatment over placebo group. On Day 4, a significant decrease in the intensity of all the symptoms (headache, tiredness, ear ache, sleeplessness, sore throat, nasal secretion, phlegm, frequency and intensity of cough) was observed for *A. paniculata* group [90].

Liver enzyme modulation:

Both andrographolide and 14-deoxy-11,12- didehydroandrographolide inhibited mRNA and protein expression of CYP1A2, CYP2D6, and CYP3A4 in HepG2 hepatoma cells. The lowest concentration (0.3 μm) of both diterpenoids produced more than 50% reduction in mRNA and protein expression of CYP3A4. This reduction was consistent with the enzyme activity. Both the compounds also reduced the ability of dexamethasone to induce CYP3A4 expression [91]. Andrographolide induced enhanced expression of CYP1 in PAH-responsive C57BL/6 male mice and did not alter CYP1 expression in PAH non-responsive DBA/2 male mice, ovariectomized females and orchiectomized male mice. However, the treatment with testosterone restored the effect of andrographolide on CYP1 in both orchiectomized males and ovariectomized females. This observation suggested a role for a male hormone system as a crucial mediator of modulation of CYP1 expression by andrographolide [92]. Andrographolide and *A. paniculata* extract significantly (P<0.05) increased the clearance and reduced the area under concentration time curve of theophylline (1 mg/kg) in blood of male Sprague Dawley rats. The elimination half-life and mean residence time of theophylline (5 mg/kg). However, theophylline (5 mg/kg) accumulated in the blood of rats was pretreated with *A. paniculata* extract. This suggested that some herbal constituents present in *A. paniculata* extract may interact with theophylline and retard its elimination when administered at a high dose [93].

Sex hormone/function modulation:

Oral administration of the leaves extract in doses of 200, 600 and 2000 mg/kg body weight (i.e. 30, 90 and 300 fold higher than its daily therapeutic dose in humans) to pregnant rats for a period of 19 days for 200 mg/kg group and 11 days for 600 and 2000 mg/kg group respectively did not show any effect on the elevated levels of progesterone in the blood plasma of pregnant rats when compared with control groups. This suggests that *A. paniculata* at therapeutic doses cannot induce abortion [94]. Andrographolide (50 mg/kg body weight) administered to male ICR mice significantly (P<0.05) decreased the mounting latency at 120 min and 180 min and increased the mounting frequency at 180 min after treatment. This suggested an improvement in sexual functions. The pre-incubation of endothelium-intact aortic strip with andrographolide for 10 min before adding nor-epinephrine resulted in a significant reduction in nor-epinephrine effect on aortic strip tension. An observation suggested that andrographolide improves sexual function by causing smooth muscle relaxation and increasing blood flow to the penis. Also, daily treatment of male mice with andrographolide (50 mg/kg) for 2, 4, 6 or 8 weeks significantly (P<0.05) increased serum testosterone levels on week 4 and this level declined back to normal (pretreatment levels) on week 6 and 8 with continued treatment. Moreover, andrographolide (50 mg/kg) was shown to have no significant effect on sperm count and motility [95].

Toxicity:

The safety of *A. paniculata* extract (Kalmcold) in genotoxic tests has been reported. In a study, the LD50 values has been determined to be higher than 5g/kg rats body weight in an oral acute toxicity [96]. Testicular toxicity was assessed by reproductive organ weight, testicular histology and ultra-structural analysis of leydig cells. The testosterone levels were not found after 60 days treatment with ethanol extract of dried herbs of *A. paniculata* in Sprague Dawley rats at doses of 20, 200 and 1000 mg/kg which suggested the relative safe toxicity profile [97-98].

Neuroprotective:

The neuroprotective effects of andrographolide were studied on RSC96 cells in vitro. The RSC96 cell line consisting of immortalized rat Schwann cell line were treated with varying concentrations of andrographolide (0 to 50 µM), prior to the MTT assay. Cell proliferation, morphology, synthesis and nerve-specific gene expression were studied and andrographolide was found to be most effective between concentration range 0.78 and 12.5 µM. The treatment increased DNA content and promoted the gene expression of glial cell line-derived neurotrophic factor, brain-derived neurotrophic factor, ciliary neurotrophic factor, and the specific Schwann cell marker S100β (P,0.05). Andrographolide accelerated the proliferation of RSC96 cells without altering the Schwann cell phenotype [99]. In another study, andrographolide potently activated NF-E2-related factor 2 (Nrf2) and also upregulated heme oxygenase-1 (HO-1) expression in primary astrocytes. Andrographolide reduced Nrf2, ubiquitination efficiency, and turnover rate, followed by upregulation of Nrf2 mRNA between 8 and 24 h. HO-1 is a known gene target of transcription factor Nrf2, which is critically involved in cellular defense against oxidative stress [100]. Andrographolide recuperated the cognitive impairment in the social species Octodon degus (the only wild-type South American rodent that develops Alzheimer's-like pathology with age), a natural model of Alzheimer's disease. The treatment resulted in the recovery of spatial memory and learning performance, recovery of synaptic basal transmission, partial or complete protection of certain synaptic proteins and reduction of phosphorylated tau protein and amyloid beta aggregate maturation in aged degus [101]. In a similar study, andrographolide increased neural progenitor cell proliferation and the number of immature neurons in the hippocampus of 2- and 10-month-old mice compared to age-matched control mice. It also stimulated neurogenesis increasing the number of newborn dentate granule neurons. The effect of andrographolide on APPswe/PS1 \Delta E9 transgenic mouse model of Alzheimer's disease showed an increased cell proliferation and density of immature neurons in the dentate gyrus. Concomitantly the increase in neurogenesis, also induced activation of the Wnt signaling pathway in the hippocampus of wild-type and APPswe/PS1 Δ E9 mice, evident by increased levels of β -catenin, the inactive form of GSK-3 β , and NeuroD1, a Wnt target gene involved in neurogenesis [102].

Cytotoxic and anti-tumor Activity:

The inhibition of hepatoma tumor growth induced by andrographolide (10 mg/kg) was found in a xenograft mouse tumor model in vivo. The miRNA chip analysis showed an increased expression of 22 miRNAs, whereas the expression of other 10 miRNAs decreased after treatment. Functional annotation of the target genes based on the differentially expressed miRNAs suggested that the majority of the genes were involved in a variety of signaling pathways, including miRNAs in cancer, mitogen-activated protein kinases (MAPKs) and focal adhesion [103]. Yang et al [104] studied the cytotoxic effect of andrographolide on human T-ALL (T-cell acute lymphoblastic leukemia) cells. It was found that 10 µg/mL compound could significantly induce Jurkat cells' apoptosis, depending on the inhibition of PI3K/ AKT pathway. Synergistic anticancer effects of andrographolide and paclitaxel (PTX) (widely used in chemotherapy for cancer treatment) were studied against A549 NSCLC (non-small cell lung cancer) cells. Andrographolide showed a time- and concentration- dependent inhibitory effect on highly proliferative MDA-MB-231 breast cancer cell proliferation, however, the treatment did not affect normal breast epithelial cells, MCF-10A (>80 %). Increased production of reactive oxygen species (ROS) with a corresponding decrease in mitochondrial membrane potential (MMP), externalization of phosphatidylserine was observed, while the population of apoptotic cells increased with prolonged exposure to andrographolide. Additionally, caspase-3 and caspase-9 were activated while Bax and Apaf-1 expression was significantly increased with a corresponding decrease in Bcl-2 and Bcl-xL expression in andrographolide-treated cells [105]. Furthermore, andrographolide was also reported to inhibit prostate cancer cells (LNCaP, C4-2b, and PC), by targeting cell cycle regulators, CXCR3 and CXCR7 chemokine receptors [106]. 14-Deoxy-11,12didehydroandrographolide (14-DDA), a major diterpenoid of AP, induced the formation of endoplasmic reticulum (ER) vacuoles and autophagosomes, with concurrent up regulation of LC3-II in the breast carcinoma cells. The mechanism of action involved increase in cytosolic calcium concentration leading to a collapse in mitochondrial membrane potential in LC3-II cells. The ER stress pathway was significantly up regulated, DDIT3 knockdown suppressed the formation of both ER vacuoles and autophagosomes, indicating that 14-DDA-induced ER stress and autophagy is dependent on this transcription factor [107]. The inhibitory effects of andrographolide on the growth of multiple myelomas (MM) cells and its possible impact on the nuclear factor (NF)-κB signaling pathway were studied by Gao and Wang [108].

Antipyretic activity:

Antipyretic, analgesic properties of nilavembu kudineer chooranam: a classical preparation used in the treatment of chikungunya fever was reported by Anbarasu, et al [109]. Madav, et al [110] reported that andrographolide not showed any analgesic activity in hot plate test in mice while it showed significant (p < 0.05) analgesic activity in acetic acid-induced writhing in mice and Randall test in rats at 300 mg/kg dose. Authors also reported that andrographolide at 100 and 300 mg/kg, oral dose elicited significant (p < 0.05) antipyretic effect after 3 h of administration in Brewer's yeast-induced pyrexia in rats and significant (p < 0.05) anti-ulcerogenic activity in aspirin induced ulceration in rats.

Antiplatelet activity:

Phytoconstituents and extracts of *Andrographis paniculata* was reported to exhibit anti-platelet activity by various mechanism of actions viz. decreasing platelet activating factor [111] and increasing eNOS-NO/cyclic-GMP pathway by decreasing PLC2-PKC and PI3 kinase/Akt-MAPKs [112]. Inhibitory effect of *Andrographis paniculata* extract and its

active diterpenoids on platelet aggregation was studied by Thisoda, et al [113]. The results indicated that andrographolide and 14-deoxy-11,12-didehydroandrographolide significantly inhibited thrombin-induced platelet aggregation in a concentration and time-dependent manner while neoandrographolide had little or no activity. The results indicated that the standardized Andrographis paniculata extract may contain other anti-platelet compounds, which contribute to high anti-platelet activity. Amroyan, et al [114] tested andrographolide for PAF-induced platelet aggregation, where, andrographolide inhibited PAF-induced human blood platelet aggregation in a dose dependent manner (IC50 \sim 5 μ M). These results indicated that andrographolide has a mechanism of action different from that of non-steroidal anti-inflammatory drugs (NSAID) and most likely associated with the cardiovascular and antithrombotic activity described of Andrographis paniculata. Wu, et al [115] isolated two new flavones designated as andropaniculosin A and ropaniculoside A and 30 known compounds from the whole plants of Andrographis paniculata.

Adverse effects:

An overdose of *Andrographis paniculata* extracts caused vomiting, gastric discomfort and loss of appetite that may be due to very high bitter taste of the herb [116]. Though this plant or its extract is safe, it is not to be taken during pregnancy as it is classified under class 2b in botanical safety hand book [117].

CONCLUSION

Andrographis paniculata has been used to cure a variety of disorders, including liver damage, infection, hyperglycemia, cancer, and other ailments. Andrographolide is a diterpenoid lactone with a variety of pharmacological properties as defined by indigenous medicine. In addition to its many medicinal use, andrographolide has some adverse effects such as nausea, vomiting, and loss of appetite that can only be detected when overdose. As a result, researchers may go on to build potent formulations containing Andrographis paniculata and its isolated molecule, andrographolide, using herbal drug delivery vehicles. For a variety of causes, human invading microorganisms are becoming resistant to available medicines. Because A. paniculata acts on immunological modulation, medication resistance is less likely to occur. Even though A. paniculata has potential antibacterial activity, more research on the method of action, impact of available antimicrobial drugs, and specific administration route and schedule is needed. The active ingredients of A. paniculata could be a source of antibacterial agents, and investigating their therapeutic potential based on clinical implications is worthwhile. We have investigated significant antimicrobial compounds in A. paniculata, but little is known about their molecular pathways in response to bacteria or host-infected cells. Human-invading germs are growing resistant to existing medications due to a variety of factors. Medication resistance is less likely since A. paniculata works on immunological regulation. Despite the fact that A. paniculata has potential antibacterial activity, more research on the mechanism of action, the impact of current antimicrobial medicines, and the specific administration route and schedule is required. The active components of A. paniculata could be a source of antibacterial agents, and it is worthwhile to investigate their therapeutic potential based on clinical implications. We found antimicrobial chemicals in A. paniculata, but little is known about their molecular processes in response to bacteria or host-infected cells. If we consider the overall efficacy of A. paniculata treatment, it would be a worthy consideration as a natural product treatment option for acute URTIs as currently, there is a lack of compelling therapeutic opportunity for IDs.

REFERENCES

- 1. Mishra SK, Sangwan NS, Sangwan RS. *Andrographis paniculata* (Kalmegh): A review, Pharmacognosy Reviews, 2007; 1:283-298.
- 2. Kabeeruddin M, Kitabul Advia, 2, Aligarh Barqi Press, Delhi, 1937, 148-150.
- 3. Shahid A, *Andrographis paniculata*: A review of pharmacological activities and clinical effects, Alternative Medicine Review, 2011; 16:66-77.
- 4. Abhishek N et al. Biological activities of Kalmegh (Andrographis paniculata Nees.) and its active principles—A review. 2010; 1(2):125-135.
- 5. Dua VK, Ojha VP, Roy R, Joshi BC, Valecha N, Devi CU *et al*, Anti-malarial activity of some xanthones isolated from the roots of *Andrographis paniculata*, J.Ethnopharmacol, 2004; 95:247-251.
- 6. Namita, P.; Mukesh, R. Medicinal plants used as antimicrobial agents: A review. Int. Res. J. Pharm. 2012, 3, 31-40.
- 7. Levy, S.B.; Marshall, B. Antibacterial resistance worldwide: Causes, challenges and responses. Nat. Med. **2004**, 10, S122–S129.
- 8. Thomas, M.; Bomar, P.A. Upper Respiratory Tract Infection. In StatPearls; StatPearls Publishing: Treasure Island, FL, USA, 2020.
- 9. Aabenhus, R.; Hansen, M.P.; Saust, L.T.; Bjerrum, L. Characterisation of antibiotic prescriptions for acute respiratory tract infections in Danish general practice: A retrospective registry based cohort study. NPJ Prim. Care Respir Med. **2017**, 27, 37.
- 10. O'Connor, R.; O'Doherty, J.; O'Regan, A.; Dunne, C. Antibiotic use for acute respiratory tract infections (ARTI) in primary care; what factors affect prescribing and why is it important? A narrative review. Ir. J. Med. Sci. **2018**, 187, 969–986. [PubMed]
- 11. O'Doherty, J.; Leader, L.F.W.; O'Regan, A.; Dunne, C.; Puthoopparambil, S.J.; O'Connor, R. Over prescribing of antibiotics for acute respiratory tract infections; a qualitative study to explore Irish general practitioners' perspectives. BMC Fam. Pract. **2019**, 20, 27.

- 12. Suriyo, T.; Pholphana, N.; Ungtrakul, T.; Rangkadilok, N.; Panomvana, D.; Thiantanawat, A.; Pongpun, W.; Satayavivad, J.Clinical Parameters following Multiple Oral Dose Administration of a Standardized Andrographis paniculata Capsule in Healthy
- Thai Subjects. Planta Med. **2017**, 83, 778–789. [CrossRef]
- 13. Poolsup, N.; Suthisisang, C.; Prathanturarug, S.; Asawamekin, A.; Chanchareon, U. Andrographis paniculata in the symptomatic treatment of uncomplicated upper respiratory tract infection: Systematic review of randomized controlled trials. J. Clin. Pharm.
- Ther. **2004**, 29, 37–45. [CrossRef]
- 14. Little, A. Review: Antibiotics are not effective for upper respiratory tract infection in children. Evid. Based Nurs. **1999**, 2, 77.
- 15. DeCorte, B.L. Underexplored Opportunities for Natural Products in Drug Discovery. J. Med. Chem. **2016**, 59, 9295–9304.
- 16. Fang, J.; Cai, C.; Wang, Q.; Lin, P.; Zhao, Z.; Cheng, F. Systems Pharmacology-Based Discovery of Natural Products for Precision Oncology Through Targeting Cancer Mutated Genes. CPT Pharmacomet. Syst. Pharmacol. **2017**, 6. 177–187.
- 17. Hazrati, S.; Govahi, M.; Sedaghat, M.; Kashkooli, A.B. A comparative study of essential oil profile, antibacterial and antioxidant activities of two cultivated Ziziphora species (Z. clinopodioides and Z. tenuior). Ind. Crop. Prod. **2020**, 157, 7. [CrossRef]
- 18. De Veras, B.O.; de Oliveira, J.R.S.; de Menezes Lima, V.L.; do Amaral Ferraz Navarro, D.M.; de Oliveira Farias de Aguiar, J.C.R.; de Medeiros Moura, G.M.; da Silva, J.W.; de Assis, C.R.D.; Gorlach-Lira, K.; de Assis, P.A.C.; et al. The essential oil of the leaves
- of Verbesina macrophylla (Cass.) S.F.Blake has antimicrobial, anti-inflammatory and antipyretic activities and is toxicologically safe. J. Ethnopharmacol. **2021**, 265, 113248. [CrossRef]
- 19. de Araujo, M.R.C.; Maciel, P.P.; Castellano, L.R.C.; Bonan, P.R.F.; Alves, D.D.N.; de Medeiros, A.C.D.; de Castro, R.D. Efficacy of essential oil of cinnamon for the treatment of oral candidiasis: A randomized trial. Spec. Care Dent. **2021**, 9. [CrossRef]
- 20. Freires, I.A.; Denny, C.; Benso, B.; de Alencar, S.M.; Rosalen, P.L. Antibacterial Activity of Essential Oils and Their Isolated Constituents against Cariogenic Bacteria: A Systematic Review. Molecules **2015**, 20, 7329–7358. [CrossRef] [PubMed]
- 21. Rodriguez-Garcia, A.; Hosseini, S.; Martinez-Chapa, S.O.; Cordell, G.A. Multi-target Activities of Selected Alkaloids and Terpenoids. Mini Rev. Org. Chem. **2017**, 14, 272–279. [CrossRef]
- 22. Rodrigues, T.; Reker, D.; Schneider, P.; Schneider, G. Counting on natural products for drug design. Nat. Chem. **2016**, 8, 531–541.[CrossRef] [PubMed]
- 23. Keiser, M.J.; Setola, V.; Irwin, J.J.; Laggner, C.; Abbas, A.I.; Hufeisen, S.J.; Jensen, N.H.; Kuijer, M.B.; Matos, R.C.; Tran, T.B.; et al. Predicting new molecular targets for known drugs. Nature **2009**, 462, 175–181. [CrossRef] [PubMed]
- 24. Xie, L.; Xie, L.; Kinnings, S.L.; Bourne, P.E. Novel computational approaches to polypharmacology as a means to define responses to individual drugs. Annu. Rev. Pharmacol. Toxicol. **2012**, 52, 361–379. [CrossRef]
- 25. Yildirim, M.A.; Goh, K.I.; Cusick, M.E.; Barabasi, A.L.; Vidal, M. Drug-target network. Nat. Biotechnol. **2007**, 25, 1119–1126.[CrossRef]
- 26. Hopkins, A.L. Network pharmacology: The next paradigm in drug discovery. Nat. Chem. Biol. **2008**, 4, 682–690. [CrossRef] [PubMed]
- 27. Reddy, A.S.; Tan, Z.; Zhang, S. Curation and analysis of multitargeting agents for polypharmacological modeling. J. Chem. Inf. Model **2014**, 54, 2536–2543. [CrossRef] [PubMed]
- 28. Reddy, A.S.; Zhang, S. Polypharmacology: Drug discovery for the future. Expert Rev. Clin. Pharmacol. **2013**, 6, 41–47. [CrossRef] [PubMed]
- 29. Taylor, W.F.; Yanez, M.; Moghadam, S.E.; Moridi Farimani, M.; Soroury, S.; Ebrahimi, S.N.; Tabefam, M.; Jabbarzadeh, E. 7-epi- Clusianone, a Multi-Targeting Natural Product with Potential Chemotherapeutic, Immune-Modulating, and Anti-Angiogenic
- Properties. Molecules 2019, 24, 4415. [CrossRef] [PubMed]
- 30. Li, J.W.; Vederas, J.C. Drug discovery and natural products: End of an era or an endless frontier? Science **2009**, 325, 161–165.[CrossRef]
- 31. Ismail, N.A.; Hossain, M.S.; Mustafa, N.H.M.; Phang, I.C. Morpho-physiological characterizatics, selected macronutrient uptak, and oxidative stress level of Andrographis paniculata under salinity condition. J. Teknol. **2015**, 77, 135–140. [CrossRef]
- 32. Hossain, M.S.; Urbi, Z.; Evamoni, F.Z.; Zohora, F.T.; Rahman, K.M.H. A secondary research on medicinal plants mentioned in the Holy Qur'an. J. Med. Plants **2016**, 15, 81–97.
- 33. Urbi, Z.; Hossain, M.S.; Rahman, K.M.H.; Zayed, T.M. Grape: AMedicinal Fruit Species in the Holy Qur'an and its Ethnomedinical Importance Department of Basic Medical Sciences, Faculty of Pharmacy. World Appl. Sci. J. **2014**, 30, 253–265. [CrossRef]
- 34. Patridge, E.; Gareiss, P.; Kinch, M.S.; Hoyer, D. An analysis of FDA-approved drugs: Natural products and their derivatives. Drug Discov. Today **2016**, 21, 204–207. [CrossRef]

- 35. Kumar, A.; Dora, J.; Singh, A.; Tripathi, R. A review on King of Bitter (Kalmegh). Int. J. Res. Pharm. Chem. **2012**, 2, 116–124.
- 36. Akbar, S. Andrographis paniculata: A review of pharmacological activities and clinical effects. Altern. Med. Rev. **2011**, 16, 66–77.
- 37. Benoy, G.K.; Animesh, D.K.; Aninda, M.; Priyanka, D.K.; Sandip, H. An overview on Andrographis paniculata (Burm. F.) Nees. Int. J. Res. Ayur. Pharm. **2012**, 3, 752–760.
- 38. Hossain, M.S.; Urbi, Z. Effect of Naphthalene Acetic Acid on the Adventitious Rooting in Shoot Cuttings of Andrographis paniculata (Burm.f.) Wall. ex Nees: An Important Therapeutical Herb. Int. J. Agron. **2016**, 2016, 1–6. [CrossRef]
- 39. Hossain, M.S.; Urbi, Z.; Sule, A.; Hafizur Rahman, K.M. Andrographis paniculata (Burm. f.) Wall. ex Nees: A review of ethnobotany, phytochemistry, and pharmacology. Sci. World J. **2014**, 2014, 274905. [CrossRef] [PubMed]
- 40. Anju, D.; Jugnu, G.; Kavita, S.; Arun, N.; Sandeep, D. A review on medicinal prospectives of Andrographis paniculata Nees. J. Pharm. Sci. Innov. **2012**, 1, 1–4.
- 41. Mussard, E.; Jousselin, S.; Cesaro, A.; Legrain, B.; Lespessailles, E.; Esteve, E.; Berteina-Raboin, S.; Toumi, H. Andrographis paniculata and Its Bioactive Diterpenoids Against Inflammation and Oxidative Stress in Keratinocytes. Antioxidants **2020**, 9, 530. [CrossRef]
- 42. Lee, D.; Baek, C.Y.; Hwang, J.H.; Kim, M.Y. Andrographis paniculata Extract Relieves Pain and Inflammation in Monosodium Iodoacetate-Induced Osteoarthritis and Acetic Acid-Induced Writhing in Animal Models. Processes **2020**, 8, 873. [CrossRef]
- 43. Li, X.; Yuan, K.; Zhu, Q.; Lu, Q.; Jiang, H.; Zhu, M.; Huang, G.; Xu, A. Andrographolide Ameliorates Rheumatoid Arthritis by Regulating the Apoptosis-NETosis Balance of Neutrophils. Int. J. Mol. Sci. **2019**, 20, 5035. [CrossRef]
- 44. Gu, L.; Yu, Q.; Li, Q.; Zhang, L.; Lu, H.; Zhang, X. Andrographolide Protects PC12 Cells against _-Amyloid-Induced Autophagy- Associated Cell Death Through Activation of the Nrf2-Mediated p62 Signaling Pathway. Int. J. Mol. Sci. **2018**, 19, 2844. [CrossRef] [PubMed]
- 45. Mussard, E.; Cesaro, A.; Lespessailles, E.; Legrain, B.; Berteina-Raboin, S.; Toumi, H. Andrographolide, a Natural Antioxidant: An Update. Antioxidants **2019**, 8, 571. [CrossRef] [PubMed]
- 46. Akhtar, M.T.; Bin Mohd Sarib, M.S.; Ismail, I.S.; Abas, F.; Ismail, A.; Lajis, N.H.; Shaari, K. Anti-Diabetic Activity and Metabolic Changes Induced by Andrographis paniculata Plant Extract in Obese Diabetic Rats. Molecules **2016**, 21, 1026. [CrossRef]
- 47. Qader, S.W.; Abdulla, M.A.; Chua, L.S.; Najim, N.; Zain, M.M.; Hamdan, S. Antioxidant, total phenolic content and cytotoxicity evaluation of selected Malaysian plants. Molecules **2011**, 16, 3433–3443. [CrossRef] [PubMed]
- 48. Liu, Y.T.; Chen, H.W.; Lii, C.K.; Jhuang, J.H.; Huang, C.S.; Li, M.L.; Yao, H.T. A Diterpenoid, 14-Deoxy-11, 12-Didehydroandrographolide, in Andrographis paniculata Reduces Steatohepatitis and Liver Injury in Mice Fed a High-Fat and High-Cholesterol Diet.

Nutrients **2020**, 12, 523. [CrossRef] [PubMed]

- 49. Ismail, S.; Hanapi, N.A.; Ab Halim, M.R.; Uchaipichat, V.; Mackenzie, P.I. Effects of Andrographis paniculata and Orthosiphon stamineus extracts on the glucuronidation of 4-methylumbelliferone in human UGT isoforms. Molecules **2010**, 15, 3578–3592. [CrossRef]
- 50. Loh, S.H.; Tsai, Y.T.; Huang, S.F.; Yu, T.C.; Kuo, P.C.; Chao, S.C.; Chou, M.F.; Tsai, C.S.; Lee, S.P. Effects of Andrographolide on Intracellular pH Regulation, Cellular Migration, and Apoptosis in Human Cervical Cancer Cells dagger. Cancers **2020**, 12, 387.[CrossRef]
- 51. Panossian, A.; Brendler, T. The Role of Adaptogens in Prophylaxis and Treatment of Viral Respiratory Infections. Pharmaceuticals **2020**, 13, 236. [CrossRef]
- 52. Wang, Y.; Jiao, J.; Yang, Y.; Yang, M.; Zheng, Q. Screening and Identification for Immunological Active Components from Andrographis Herba Using Macrophage Biospecific Extraction Coupled with UPLC/Q-TOF-MS. Molecules **2018**, 23, 1047.[CrossRef]
- 53. Kaur, R.; Sharma, P.; Gupta, G.K.; Ntie-Kang, F.; Kumar, D. Structure-Activity-Relationship and Mechanistic Insights for Anti-HIV Natural Products. Molecules **2020**, 25, 2070. [CrossRef]
- 54. Calabrese, C.; Berman, S.H.; Babish, J.G.; Ma, X.; Shinto, L.; Dorr, M.; Wells, K.; Wenner, C.A.; Standish, L.J. A phase I trial of andrographolide in HIV positive patients and normal volunteers. Phytother. Res. **2000**, 14, 333–338. [CrossRef]
- 55. Chuthaputti, A.; Pornpatkul, V.; Suwankiri, U. The Efficacy of Andrographis paniculata (Burm. f.) Wall. ex Nees for the Relief of the Symptoms of Influenza. J. Thai Tradit. Altern. Med. **2007**, 5, 1–10.
- 56. Hancke, J.; Burgos, R.; Caceres, D.; Wikman, G. A double-blind study with a new monodrug Kan Jang: Decrease of symptoms and improvement in the recovery from common colds. Phytother. Res. **1995**, 9, 559–562. [CrossRef]
- 57. Kulichenko, L.L.; Kireyeva, L.V.; Malyshkina, E.N.; Wikman, G. A randomized, controlled study of Kan Jang versus amantadine in the treatment of influenza in Volgograd. J. Herb. Pharmacother. **2003**, 3, 77–93. [CrossRef] [PubMed]
- 58. Melchior, J.; Palm, S.; Wikman, G. Controlled clinical study of standardized Andrographis paniculata extract in common cold—A pilot trial. Phytomedicine **1997**, 3, 315–318. [CrossRef]
- 59. Saxena, R.C.; Singh, R.; Kumar, P.; Yadav, S.C.; Negi, M.P.; Saxena, V.S.; Joshua, A.J.; Vijayabalaji, V.; Goudar, K.S.; Venkateshwarlu, K.; et al. A randomized double blind placebo controlled clinical evaluation of extract of

- Andrographis paniculata (KalmCold) in patients with uncomplicated upper respiratory tract infection. Phytomedicine **2010**, 17, 178–185. [CrossRef]
- 60. Thamlikitkul, V.; Dechatiwongse, T.; Theerapong, S.; Chantrakul, C.; Boonroj, P.; Punkrut, W.; Ekpalakorn, W.; Boontaeng, N.; Taechaiya, S.; Petcharoen, S.; et al. Efficacy of Andrographis paniculata, Nees for pharyngotonsillitis in adults. J. Med. Assoc. Thai

1991, 74, 437–442. [PubMed]

- 61. Widjajakusuma, E.C.; Jonosewojo, A.; Hendriati, L.; Wijaya, S.; Surjadhana, A.; Sastrowardoyo, W.; Monita, N.; Muna, N.M.; Fajarwati, R.P.; Ervina, M.; et al. Phytochemical screening and preliminary clinical trials of the aqueous extract mixture of Andrographis paniculata (Burm. f.) Wall. ex Nees and Syzygium polyanthum (Wight.) Walp leaves in metformin treated patients with type 2 diabetes. Phytomedicine **2019**, 55, 137–147. [CrossRef]
- 62. Tang, T.; Targan, S.R.; Li, Z.S.; Xu, C.; Byers, V.S.; Sandborn, W.J. Randomised clinical trial: Herbal extract HMPL-004 in active ulcerative colitis-a double-blind comparison with sustained release mesalazine. Aliment. Pharmacol. Ther. **2011**, 33, 194–202.

[CrossRef]

- 63. Sandborn, W.J.; Targan, S.R.; Byers, V.S.; Rutty, D.A.; Mu, H.; Zhang, X.; Tang, T. Andrographis paniculata extract (HMPL-004) for active ulcerative colitis. Am. J. Gastroenterol. **2013**, 108, 90–98. [CrossRef]
- 64. Phunikhom, K.; Khampitak, K.; Aromdee, C.; Arkaravichien, T.; Sattayasai, J. Effect of Andrographis paniculata Extract on Triglyceride Levels of the Patients with Hypertriglyceridemia: A Randomized Controlled Trial. J. Med. Assoc. Thai **2015**, 98 (Suppl.
- 6), S41-S47.
- 65. Islam, M.T.; Ali, E.S.; Uddin, S.J.; Islam, M.A.; Shaw, S.; Khan, I.N.; Saravi, S.S.S.; Ahmad, S.; Rehman, S.; Gupta, V.K.; et al. Andrographolide, a diterpene lactone from Andrographis paniculata and its therapeutic promises in cancer. Cancer Lett. 2018,
- 420, 129-145. [CrossRef]
- 66. Hancke, J.L.; Srivastav, S.; Caceres, D.D.; Burgos, R.A. A double-blind, randomized, placebo-controlled study to assess the efficacy of Andrographis paniculata standardized extract (ParActin(R)) on pain reduction in subjects with knee osteoarthritis. Phytother.

Res. 2019, 33, 1469–1479. [CrossRef] [PubMed]

- 67. Bertoglio, J.C.; Baumgartner, M.; Palma, R.; Ciampi, E.; Carcamo, C.; Caceres, D.D.; Acosta-Jamett, G.; Hancke, J.L.; Burgos, R.A. Andrographis paniculata decreases fatigue in patients with relapsing-remitting multiple sclerosis: A 12-month double-blind
- placebo-controlled pilot study. BMC Neurol. 2016, 16, 77. [CrossRef] [PubMed]
- 68. Barth, A.; Hovhannisyan, A.; Jamalyan, K.; Narimanyan, M. Antitussive effect of a fixed combination of Justicia adhatoda, Echinacea purpurea and Eleutherococcus senticosus extracts in patients with acute upper respiratory tract infection: A comparative,
- randomized, double-blind, placebo-controlled study. Phytomedicine 2015, 22, 1195–1200. [CrossRef]
- 69. Sgorlon, S.; Colitti, M.; Asquini, E.; Ferrarini, A.; Pallavicini, A.; Stefanon, B. Administration of botanicals with the diet regulates gene expression in peripheral blood cells of Sarda sheep during ACTH challenge. Domest Anim. Endocrinol. **2012**, 43, 213–226.

[CrossRef]

- 70. Raghavan, R.; Cheriyamundath, S.; Madassery, J. Andrographolide, a new potential NF-kappa B inhibitor: Docking simulation and evaluation of drug-likeness. Mol. Simulat. **2012**, 38, 582–588. [CrossRef]
- 71. Novianto, F.; Zulkarnain, Z.; Mana, T.A. A Clinical Observation to Understand the Safety of Herbs Used for Diabetes mellitus. Media Penelit Pengem **2018**, 28, 9–14. [CrossRef]
- 72. He, Y.; Yang, J.; Zeng, G.; Shen, T.; Fontaine, R.E.; Zhang, L.; Shi, G.; Wang, Y.; Li, Q.; Long, J. Risk factors for critical disease and death from hand, foot and mouth disease. Pediatr. Infect. Dis. J. **2014**, 33, 966–970. [CrossRef] [PubMed]
- 73. Ciampi, E.; Uribe-San-Martin, R.; Carcamo, C.; Cruz, J.P.; Reyes, A.; Reyes, D.; Pinto, C.; Vasquez, M.; Burgos, R.A.; Hancke, J. Efficacy of andrographolide in not active progressive multiple sclerosis: A prospective exploratory double-blind, parallel-group, randomized, placebo-controlled trial. BMC Neurol. **2020**, 20, 173. [CrossRef]
- 74. Panossian, A.; Hovhannisyan, A.; Mamikonyan, G.; Abrahamian, H.; Hambardzumyan, E.; Gabrielian, E.; Goukasova, G.; Wikman, G.; Wagner, H. Pharmacokinetic and oral bioavailability of andrographolide from Andrographis paniculata fixed combination Kan Jang in rats and human. Phytomedicine **2000**, 7, 351–364. [CrossRef]
- 75. Li, L.; Wijaya, H.; Samanta, S.; Lam, Y.; Yao, S.Q. In situ imaging and proteome profiling indicate andrographolide is a highly promiscuous compound. Sci. Rep. **2015**, 5, 11522. [CrossRef] [PubMed]
- 76. Bisson, J.; McAlpine, J.B.; Friesen, J.B.; Chen, S.N.; Graham, J.; Pauli, G.F. Can Invalid Bioactives Undermine Natural Product- Based Drug Discovery? J. Med. Chem. **2016**, 59, 1671–1690. [CrossRef] [PubMed]
- 77. Singh PK, Roy S and Dey S. Antimicrobial activity of Andrographis paniculata. Fitoterapia. 74; 2003: 692-694.
- 78. Chandrasekaran CV, Gupta A and Agarwal A. Effect of an extract of *Andrographis paniculata* leaves on inflammatory and allergic mediators *in vitro*. Journal of Ethnopharmacology. 129; 2010: 203-207.

- 79. Chandrasekaran CV, Thiyagarajan P, Deepak HB and Agarwal A. *In vitro* modulation of LPS/calcimycin induced inflammatory and allergic mediators by pure compounds of *Andrographis paniculata* (King of bitters) extract. International Immunopharmacology. 11; 2011: 70-84.
- 80. Xu C, Chou GX, Wang CH and Wang ZT. Rare noriridoids from the roots of *Andrographis paniculata*. Phytochemistry. **77**; 2012: 275- 279.
- 81. Parichatikanond W, Suthisisang C, Dhepakson P and Herunsalee A.
- Study of anti-inflammatory activities of the pure compounds from *Andrographis paniculata* (Burm.f.) Nees and their effects on gene expression. International Immunopharmacology. 10; 2010: 1361-1373.
- 82. Liu J, Wang ZT and Ge BX. Andrograpanin, isolated from *Andrographis paniculata*, exhibits anti-inflammatory property in lipopolysaccharide-induced macrophage cells through down-regulating the p38 MAKs signaling pathways. International Immunopharmacology. 8; 2008: 951-958.
- 83. Neogy S, Das S, Mahapatra SK, Mandal N and Roy S. Amelioratory effect of *Andrographis paniculata* Nees on liver, kidney, heart, lung and spleen during nicotine induced oxidative stress. Environmental Toxicology and Pharmacology. 25; 2008: 321-328.
- 84. Akowuah GA, Zhari I, Norhayati I and Mariam A. HPLC and HPTLC densitometric determination of Andrographolides and antioxidant potential of *Andrographis paniculata*. Journal of Food Compostion and Analysis. 19; 2006: 118-126.
- 85. Akowuah GA, Zhari I, Mariam A and Yam MF. Absorption of andrographolides from *Andrographis paniculata* and its effect on CCl4-induced oxidative stress in rats. Food and Chemical Toxicology. 47; 2009: 2321-2326.
- 86. Dua VK, Oha VP, Biswas S, Valecha N, Singh N and Sharma VP. Antimalarial activity of different fractions isolated from the leaves of *Andrographis paniculata*. Journal of Medicinal and Aromatic Plant Sciences. 21; 1999: 1069-1073.
- 87. Misra P, Pal NL, Guru PY, Kariya JC, Srivastava V and Tandon JC. Antimalarial activity of *Andrographis paniculata* (Kamelgh) against *Plasmodium berghei* NK65 in Mastomys natalensis. International Journal of Pharmceutics. 30: 1992: 263-274.
- 88. Govindarajan M. Evaluation of *Andrographis paniculata* Burm. f. (Family: Acanthaceae) extracts against *Culex quinquefasciatus* (Say) and *Aedes aegypti* (Linn). Asian Pacific Journal of Tropical Medicine. 4; 2011: 176-181.
- 89. Saxena RC, Singh R, Kumar P, Yadav SC, Neigh MP and Saxena VS. A randomized double blind placebo controlled clinical evaluation of extract of *Andrographis paniculata* (Kalmcold) in patients with uncomplicated upper respiratory tract infection. Phytomedicine. 17; 2010: 178-185.
- 90. Caceres DD, Hancke JL, Burgos RA, Sandberg F and Wikman GK. Use of visual analogue scale measurement (VAS) to assess the effectiveness of standardized *Andrographis paniculata* extract SHA-10 in reducing the symptoms of common cold. A randomized double blind-placebo study. Phytomedicine. 6; 1999: 217-223.
- 91. Obi JP, Suleiman SF, Muhammad TS and Tan ML. Andrographolide and 14-deoxy-11, 12-didehydroandrographolide inhibits cytochrome P450s in HepG2 hepatoma cells. Life Sciences. 88; 2001: 447-454.
- 92. Imsanguan P, Pongamphai S, Douglas S and Teppaitoon W. Supercritical antisolvent precipitation of andrographolide from *Andrographis paniculata* extracts: effect of pressure, temperature and CO2 flow rate. Powder Technology. 200; 2010: 246-253.
- 93. Chien CF, Wu YT, Lee WC, Lin, LC and Tsai TH. Herb drug interaction of *Andrographis paniculata* extract and andrographolide on the pharmacokinetics of theophylline in rats. Chemico-Biological Interactions. 184; 2010: 458-465.
- 94. Panossian A, Kochikian A, Gabrielian E, Muradian R, Stephanian H and Armenian F. Effect of *Andrographis paniculata* extract on progesterone in blood plasma of pregnant rats. Phytomedicine. 6; 1999: 157-161.
- 95. Sattayasai J, Srisuwan S, Arkaravichien T and Aromedee C. Effects of andrographolide on sexual functions, vascular reactivity and serum testosterone level in rodents. Food and Chemical Toxicology. 48; 2010: 1934-1938.
- 96. Chandrasekaran CV, Thiyagarajan P, Sundarajan K, Goudar KS, Deepak M and Murali B. Evaluation of the genotoxic potential and acute oral toxicity of standardized extract of *Andrographis paniculata* (Kalmcold). Food and Chemical Toxicology. 47; 2009: 1892-1902.
- 97. Burgos RA, Caballero EE, Sanchez NS, Schroeder RA, Wikman GK and Hancke JL. Testicular toxicity assessment of *Andrographis paniculata* dried extract in rats. Journal of Ethnopharmacology. 58; 1997: 219-224.
- 98. Balu S, Alagesaboopathi C and Elango V. Antipyretic activities of some species of *Andrographis* Wall. Ancient Science Life. 12; 1992: 399-402.
- 99. Xu F, Wu H, Zhang K, Zheng L, Zhao J. Proneurogenic effects of andrographolide on RSC96 Schwann cells *in vitro*. Molecular Medicine Reports. 14; 2016: 3573-3580.
- 100. Wong SY, Tan MG, Wong PT, Herr DR and Lai MK. Andrographolide induces Nrf2 and heme oxygenase 1 in astrocytes by activating p38 MAPK and ERK. Journal of Neuroinflammation. 13; 2016: 251.
- 101. Rivera DS, Lindsay C, Codocedo JF, Morel I, Pinto C, Cisternas P, Bozinovic F and Inestrosa NC. Andrographolide recovers cognitive impairment in a natural model of Alzheimer's disease (Octodon degus). Neurobiology of Aging. 46; 2016: 204-220.
- 102. Varela-Nallar, L, Arredondo SB, Tapia-Rojas C, Hancke J and Inestrosa NC. Andrographolide Stimulates Neurogenesis in the Adult Hippocampus. Neural Plast. 2015. doi: 10.1155/2015/935403.
- 103. Lu B, Lu B, Sheng Y, Zhang J and Zheng Z. The altered microRNA profile in andrographolide-induced inhibition of hepatoma tumour growth. Gene. 588; 2016: 124-133.

- 104. Yang T, Yao S, Zhang X and Guo Y. Andrographolide inhibits growth of human T-cell acute lymphoblastic leukemia Jurkat cells by downregulation of PI3K/AKT and upregulation of p38 MAPK pathways. Drug Design Development and Therapy. 11; 2016: 1389-1397.
- 105. Banerjee M, Chattopadhyay S, Choudhuri T, Bera R, Kumar S, Chakraborty B and Mukherjee SK. Cytotoxicity and cell cycle arrest induced by andrographolide lead to programmed cell death of MDAMB-231 breast cancer cell line. Journal of Biomedical Science. 23; 2016: 40-56.
- 106. Mir H, Kapur N, Singh R, Sonpavde G, Lillard JW and Singh S. Andrographolide inhibits prostate cancer by targeting cell cycle regulators, CXCR3 and CXCR7 chemokine receptors. Cell Cycle 15; 2016: 819-826.
- 107. Tan HK, Muhammad TST and Tan ML. 14-Deoxy-11,12-didehydroandrographolide induces DDIT3-dependent endoplasmic reticulum stress mediated autophagy in T-47D breast carcinoma cells. Toxicology and Applied Pharmacology. 300; 2016: 55-69.
- 108. Gao H and Wang J. Andrographolide inhibits multiple myeloma cells by inhibiting the TLR4/NF-κB signaling pathway. Molecular Medicine Reports. 13; 2016: 1827-1832.
- 109. Anbarasu K, Manisenthil KT and Ramachandran S. 2014. Antipyretic, anti-inflammatory and analgesic properties of nilavembu kudineer choornam: a classical preparation used in the treatment of chikungunya fever. Asian Pacific Journal of Tropical Medicine. 10; 2016: 819-823.
- 110. Madav S, Pathihi HC, Mishra, SK. Analgesic, Antipyretic and Anti-ulcerogenic effects of Andrographolide. Indian Journal of Pharmaceutical Sciences. 57(3); 1995: 121-125.
- 111. Lu WJ, Lee IJ and Chou DS. A novel role of andrographolide, an NF-kappa B inhibitor, on inhibition of platelet activation. The pivotal mechanisms of endothelial nitric oxide synthase/cyclic GMP. Journal of Molecular Medicine. 89; 2011: 1263-1271.
- 112. Lu WJ, Lin KH, Hsu MJ, Chou DS, Hsiao G and Shen RJ. Suppression of NK-KB signalling by and rographolid with a novel mechanism in human platelets regulatory roles of the p38 MAPK-hydroxyl radicalERK-2 cascade. Biochemical Pharmacology. 84; 2012: 914-924.
- 113. Thisoda P, Rangkadilok N, Pholphana N, Worasuttayangkurn L and Ruchirawat S. Inhibitory effect of *Andrographis paniculata* extract and its active diterpenoids on platelet aggregation. European Journal of Pharmacology. 553(1-3); 2006: 39-45.
- 114. Amroyan E, Gabrielian E, Panossian A, Wikman G and Wagner H. Inhibitory effect of andrographolide from *Andrographis paniculata* on PAF-induced platelet aggregation. Phytomedicine. 6(1); 1999: 27–31.
- 115. Wu TS, Chern HJ, Damu AG, Kuo PC, Su CR, Lee EJ and Teng CM. Flavonoids and ent-labdane iterpenoids from *Andrographis paniculata* and their antiplatelet aggregatory and vasorelaxing effects. Journal of Asian Natural Products Research. 10(1-2); 2008: 17-24.
- 116. Chang HM and But PPH. Pharmacology and applications of Chinese materia medica Singapore: World Scientific. 1986.
- 117. McGuffin M, Hobbis C, Upton R and Goldberg A. American Herbal products Association Botanical Safety Handbook: CRC Press. 1997.